ANGIOTENSIN II EXERTS ITS EFFECT ON ALDOSTERONE PRODUCTION AND POTASSIUM PERMEABILITY THROUGH RECEPTOR SUBTYPE AT₁ IN RAT ADRENAL GLOMERULOSA CELLS

GYÖRGY HAJNÓCZKY, GYÖRGY CSORDÁS, ATTILA BAGÓ, ANDREW T. CHIU* and ANDRÁS SPÄT†

Department of Physiology, Semmelweis University Medical School, H-1444 Budapest 8, P.O. Box 259, Hungary; and *Medical Products Department E.I. DuPont de Nemours and Company, Wilmington, DE 19800-0400, U.S.A.

(Received 23 July 1991; accepted 8 November 1991)

Abstract—The stimulatory effect of angiotensin II (AT) on the accumulation of inositol phosphates and on aldosterone production is abolished by the AT₁ selective receptor antagonist DuP753 while PD123177, an AT₂ antagonist, is ineffective. Similarly, a depolarizing effect of AT (inhibition of $K^+/^{86}Rb$ efflux) is prevented by DuP753. While mediators derived from phospholipase C activation have a central role in the stimulation of aldosterone production by AT, the effect of AT on K^+ permeability is mimicked neither by elevation of cytoplasmic [Ca²⁺] by ionomycin nor by kinase C activation with phorbol ester. Our results suggest that AT stimulates phospholipase C and the subsequent steroid production by glomerulosa cells through AT₁ receptors. In addition some events induced by the activation of AT₁ receptors may not be mediated by the activation of phospholipase C.

Two different binding sites of angiotensin II (AT‡) were identified recently with highly specific nonpeptide antagonists in rat adrenal cortex [1-3] and in other tissues [1, 2, 4]. Only one subtype (AT_1) was found in rabbit adrenal and liver [5], showing the possibility of tissue- and species-specific distribution of these receptor subclasses. There are few data on the messenger mechanisms associated with the two AT receptor subtypes. In liver cells only DuP753 (an antagonist of receptor subtype AT₁) inhibits the formation of inositol phosphates and activation of phosphorylase a in AT-stimulated cells [5, 6]. These data show that ligand binding to AT₁ receptors activates phospholipase C and induces a Ca²⁺ signal. The role of receptor subtype AT₂ has not been elucidated.

The response to AT is also induced by activation of phospholipase C in adrenal glomerulosa cells (for review see Refs 7 and 8). However, it is not known whether AT_1 receptors are involved in this response. We also examined the action of AT on the K^+ permeability of glomerulosa cells to elucidate whether the same AT receptor subtype and the same intracellular mediators are responsible for the effect of AT on aldosterone production and on K^+ permeability.

MATERIALS AND METHODS

Angiotensin II (Ile⁵-angiotensin II) was obtained from Serva (Heidelberg, F.R.G.), TPA from the Sigma Chemical Co. (St Louis, MO, U.S.A.), ionomycin from Calbiochem (Luzern, Switzerland)

and nonpeptide AT receptor antagonists were synthesized at DuPont (Wilmington, DE, U.S.A.). Nitrocellulose membrane filter (pore size $3 \mu m$) was from Schleicher and Schüll (Feldbach ZH, Switzerland), myo-[2- 3 H]inositol (16.6 Ci/mmol) from DuPont (Boston, MA, U.S.A.). and [86 Rb]-RbCl (0.5 Ci/g) from Izinta (Budapest, Hungary).

Rat adrenal glomerulosa cells were prepared from capsular tissues of Wistar rats (200-300 g) using collagenase and mechanical dispersion as described previously [9].

In the aldosterone experiments adrenal glomerulosa cells derived from male rats were preincubated for 3 hr in a mixture of Krebs-Ringer-bicarbonate-glucose solution and Medium 199 (2:1 v/v). The composition of Krebs-Ringer solution was modified to give the following final concentrations in mM: Na, 145; K, 3.6; Ca, 1.2 and Mg, 0.5; human serum albumin (fraction V), 2 g/L. The pH was kept at 7.4 under a mixtue of 95% $O_2 + 5\%$ CO_2 (37°). Then the cells were washed and incubated (about 10^5 cells/ $600~\mu$ L) for 60 min under identical conditions. Aldosterone content of the supernatant (after pelleting the cells) was measured by radioimmunoassay [10]. Mean basal aldosterone production was $0.89 \pm 0.28~\text{pmol}/10^5~\text{cells} \times \text{hr}$ (N = 4).

Experiments on the formation of inositol phosphates were performed as described previously [11]. Labeling period was followed by a 15-min stimulation in the presence of LiCl (10 mM). Inositol phosphates were separated by anion exchange chromatography [12].

The K⁺ permeability of the cells was examined by measuring ⁸⁶Rb efflux from isotope preloaded glomerulosa cells. Cells derived from female rats (about 10⁵) were preincubated in the presence of a

BP 43:5-G 1009

[†] Corresponding author. FAX (36) 1-1187-480.

[‡] Abbreviations: AT, angiotensin II; TPA, Arg-AVP, tetradecanoyl phorbol acetate; vasopressin.

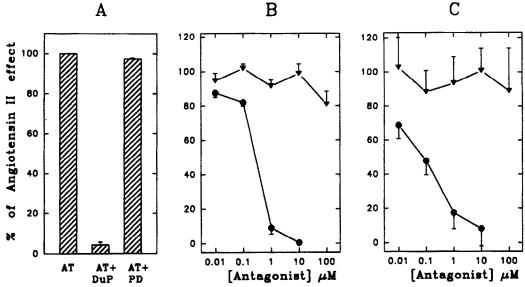


Fig. 1. Effect of selective nonpeptide AT receptor antagonists on AT-induced [³H]inositol trisphosphate accumulation (A), on AT-stimulated aldosterone production (B) and on AT-induced inhibition of ⁸⁶Rb efflux (C). Cells were stimulated with AT (25 nM) in the presence or absence of DuP753 (●) or PD123177 (▼). Means ± SEM are shown (A, N = 3; B, N = 4; C, N = 5).

tracer amount of $^{86}\text{RbCl}$ (0.2–0.5 $\mu\text{Ci}/100\,\mu\text{L}$) for 70 min at 37°. Here, the medium was buffered with N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid (20 mM) instead of bicarbonate. Isotope efflux was induced by a 15-fold dilution with isotope free medium containing stimulatory substances or solvent. Incubation was usually terminated 15 min after the onset of stimulation by vacuum filtration through nitrocellulose membrane filters. The activity of ^{86}Rb on the filter was measured by liquid scintillation counting.

RESULTS AND DISCUSSION

Maximally effective concentration of AT (25 nM) induced a 6.0 ± 0.5 -fold increase in the accumulation of [3H]inositol trisphosphates in glomerulosa cells (N = 3). This effect of AT was almost completely abolished by the AT₁-specific receptor antagonist DuP753 (10 μM) while the AT₂-specific PD123177 $(10 \,\mu\text{M})$ had a negligible effect only (Fig. 1A). The same concentration of AT induced a 14.7 ± 5.2 -fold stimulation of aldosterone production (N = 4). DuP753 dose-dependently inhibited the effect of AT (IC₅₀ approx. 250 μ M) while PD123177 did not modify the effect of AT up to $10 \,\mu\text{M}$ (Fig. 1B). Neither Dup 753 nor PD123177 exerted any significant effect on aldosterone production by control cells or cells stimulated with 8.4 mM K⁺ (data not shown). These data show that the stimulatory effect of AT on phospholipase C and on steroid production is mediated by the AT₁ receptor subtype.

A dual effect of AT was reported previously on the potassium permeability of bovine glomerulosa cells [13]. An initial increase was followed by a sustained reduction of potassium permeability. The initial increase was attributed to Ca²⁺-activated K⁺ efflux. In fact, we also found that Ca ionophores

enhanced 86Rb efflux from rat adrenal glomerulosa cells (the residual ⁸⁶Rb content after 5 min efflux was $80.6 \pm 5.9\%$ of control in the presence of $10 \,\mu\text{M}$ ionomycin, P < 0.05, N = 4). However, the Camobilizing AT (25 nM) did not induce an initial increase and evoked sustained inhibition only of ⁸⁶Rb efflux in our experiments (Fig. 2A). Therefore, it may be assumed that in AT-stimulated rat glomerulosa cells Ca2+-activated K+ efflux is masked by an unknown mechanism leading to reduced K⁺ permeability. Conversely, the reduction of K+ permeability may not be attributed to the Ca2+ signal. Considering that cytoplasmic Ca2+ exerts a mediating role in AT-induced steroid production, the idea arose that AT-evoked Ca²⁺ signal and inhibition of K+ efflux are initiated by different receptor subtypes. To test this presumption, we have examined the effect of AT antagonists also on K+ permeability. AT-induced inhibition of 86Rb efflux (5 min) was influenced by DuP753 and PD123177 in the same manner as aldosterone production (Fig. 1C).

 \dot{V}_1 -type AVP receptors in glomerulosa cells are coupled to the Ca signalling pathway [14]. Like AT, AVP exerted a significant inhibitory effect on ⁸⁶Rb efflux (Fig. 2B, P < 0.02, N = 4). (The small effect of AVP as compared to AT may be attributed to the low density of V_1 receptors [14].) The role of protein kinase C is unlikely to mediate the effect of AT or AVP on K⁺ permeability, since TPA that activates protein kinase C under similar conditions [15] was found to be completely ineffective (Fig. 2B). Thus, the effects of protein kinase C on K⁺ permeability seem to be different in bovine [16] and rat glomerulosa cells.

The present data suggest that in rat adrenal glomerulosa cells the AT₁ subtype of angiotensin receptors is coupled to Ca mobilization and thereby

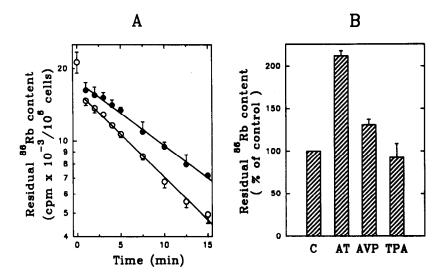


Fig. 2. Effect of AT, AVP and TPA on ⁸⁶Rb efflux. (A) Time-course of ⁸⁶Rb efflux from unstimulated (○) and AT (25 nM) treated cells (●). Fifteen-fold dilution of prelabelled cells with isotope free medium was done at 0 min. Residual ⁸⁶Rb content of the cells is shown (means ± SEM, N = 3). (B) Residual ⁸⁶Rb content of the cells after 15 min efflux in the presence of AT (25 nM), AVP (100 nM) or TPA (50 nM). Results were expressed as percentages of residual ⁸⁶Rb content of control cells (means ± SEM, N = 4-5).

to steroid production. The decrease in K+ permeability may participate in AT induced depolarization [17, 18] of the cells. This effect is also mediated by AT₁ receptors. Our results also indicate that known messengers generated through activation of phospholipase C have no role in the reduction of K⁺ permeability since the Ca ionophore ionomycin increases K⁺ permeability whereas phorbol ester, an activator of kinase C, fails to affect K⁺ permeability. These observations raise the possibility that binding of AT to AT₁ receptors, in addition to its known effects, activates a hitherto unrecognized transducing mechanism. Further experiments are required to elucidate whether the unidentified modulator of potassium channels is a G-protein coupled to the AT₁ receptor or is generated in a later step of the Ca signalling pathway.

Acknowledgements—The excellent technical help of Miss Erika Kovács and Mrs Ágnes Ribár is greatly appreciated. The aldosterone antibody was a gift from NIAMD (Bethesda, MD, U.S.A.). This work was supported by research grant OTKA No. 1111 from the Hungarian National Research Fund (OTKA) and grant No. T-176/1990 (ETT) from the Hungarian Council for Medical Sciences.

REFERENCES

- Chiu AT, Herblin WF, McCall DE, Ardecky RJ, Carini DJ, Duncia JV, Pease LJ, Wong PC, Wexler RR, Johnson AL and Timmermans PBMWM, Identification of angiotensin II receptor subtypes. Biochem Biophys Res Commun 165: 196-203, 1989.
- Whitebread S, Mele M, Kamber B and de Gasparo M, Preliminary biochemical characterization of two

- angiotensin II receptor subtypes. Biochem Biophys Res Commun 163: 284-291, 1989.
- Chiu AT, McCall DE, Aldrich PE and Timmermans PBMWM, [3H]DuP 753, a highly potent and specific radioligand for the angiotensin II-1 receptor subtype. Biochem Biophys Res Commun 172: 1195-1202, 1990.
- Rogg H, Schmid A and de Gasparo M, Identification and characterization of angiotensin II receptor subtypes in rabbit ventricular myocardium. Biochem Biophys Res Commun 173: 416-422, 1990.
- Dudley DT, Panek RL, Major TC, Lu GH, Bruns RF, Klinkefus BA, Hodges JC and Weishaar RE, Subclasses of angiotensin II binding sites and their functional significance. Mol Pharmacol 38: 370-377, 1990.
- Garcia-Sáinz AJ and Macias-Silva M, Angiotensin II stimulates phosphoinositide turnover and phosphorylase through AII-1 receptors in isolated rat hepatocytes. Biochem Biophys Res Commun 172: 780– 785, 1990.
- Spät A, Stimulus-secretion coupling in angiotensinstimulated glomerulosa cells. J Steroid Biochem 29: 443–453, 1988.
- Barret PQ, Bollag WB, Isales CM, McCarthy RT and Rasmussen H, Role of calcium in angiotensin IImediated aldosterone secretion. *Endocrine Rev* 10: 496-518, 1989.
- Spät A, Balla I, Balla T, Cragoe EJ Jr, Hajnóczky Gy and Hunyady L, Angiotensin II and potassium activate different calcium entry mechanisms in rat adrenal glomerulosa cells. J Endocrinol 122: 361-370, 1989.
- Enyedi P and Spät A, Effect of reduced extracellular sodium concentration on the function of adrenal glomerulosa cells: studies on isolated glomerulosa cells from rat. J Endocrinol 89: 417-421, 1981.
- Hajnóczky Gy, Várnai P, Holló Zs, Christensen SB, Balla T, Enyedi P and Spät A, Thapsigargin-induced increase in cytoplasmic Ca²⁺ concentration and aldosterone production in rat adrenal glomerulosa

- cells: interaction with potassium and angiotensin II. Endocrinology 128: 2639-2644, 1991.
- Berridge MJ, Dawson MRC, Downes CP, Heslop JP and Irvine RF, Changes in the levels of inositol phosphates after agonist-dependent hydrolysis of membrane phosphoinositides. *Biochem J* 212: 473-482, 1983.
- 13. Lobo MV and Marusic ET, Angiotensin II causes a dual effect on potassium permeability in adrenal glomerulosa cells. Am J Physiol 254: E144-149, 1988.
- 14. Balla T, Enyedi P, Spät A and Antoni FA, Pressortype vasopressin receptors in the adrenal cortex: properties of binding, effects on phosphoinositide metabolism and aldosterone secretion. *Endocrinology* 117: 421-423, 1985.
- Faragó A, Seprödi J and Spät A, Subcellular distribution of protein kinase C in rat adrenal glomerulosa cells. Biochem Biophys Res Commun 156: 628-633, 1988.
- Lobo MV, Mendoza RR and Marusic ET, SN-1,2 dioctanoylglycerol mimics the effects of angiotensin II on aldosterone production and potassium permeability in isolated bovine glomerulosa cells. J Steroid Biochem 35: 29-33, 1990.
- Natke E and Kabela E, Electrical responses in cat adrenal cortex: possible relation to aldosterone secretion. Am J Physiol 237: E158-162, 1979.
- Quinn SJ, Cornwall MC and Williams GH, Electrophysiological responses to angiotensin II of isolated rat adrenal glomerulosa cells. *Endocrinology* 120: 1581-1589, 1987.